

CELLULASE PRODUCTION OPTIMIZATION USING CELLULOLYTIC BACTERIA

P. Nisha*

P.G. Department of Biochemistry and Biotechnology, S.S.V. College,
Valanchirangara, Ernakulam, Kerala - 683 556, India.

ABSTRACT

Cellulase enzyme is an important enzyme used in various industries and is very expensive one. Cellulase enzyme is easy to produce from micro organisms. Optimum environmental conditions are varies for each organism for the enzyme production. This study is aimed that, maximum cellulase enzyme production from *Micrococcus* sp by changing some of the environmental parameters, enzyme activity assayed by Di Nitro Salicylic acid method with glucose as standard. The parameters studied were incubation temperature, pH, different concentrations of CMC and incubation time. It was found that, the optimum parameters for higher production of Cellulase enzyme was at 37°C of incubation temperature, pH 8, 1.5% of CMC and 72 hr of incubation time, 0.9490 IU/L.

Keywords: Cellulase, *Micrococcus* sp, DNSA, CMC, Enzyme assay.

INTRODUCTION

Cellulase is an enzyme capable to degrade cellulose in the environment. Cellulases degrade cellulose by the enzyme activities such as endoglucanase, which is known as carboxymethyl cellulase (CMCase) (endo-1,4- β -D-glucanase, EG, EC 3.2.1.4), exoglucanase (known as cellobiohydrolase) (exo-1,4- β -D-glucanase, CBH, EC 3.2.1.91); and β -glucosidase (1,4- β -D-glucosidase, BG, EC 3.2.1.21)^{1,2}. Many micro organisms such as bacteria, fungi, actinomycetes and yeasts are capable of producing extracellular cellulase enzyme³. Number of studies on cellulase enzyme production have been concentrate on fungi than with bacterial strains.^{4,5,6,7} Cellulase producing bacteria were isolated from various habitats like soil, hot springs, organic matters, faeces of ruminants decayed plant materials, and composts to obtain effective enzyme producer⁸. Production of cellulase enzyme is very much dependent on Culture and environmental parameters⁹ such as temperature, carbon sources, aeration, incubation time, medium ingredients, pH of the medium and cellulose quality¹⁰.

Cellulase is a complex enzyme and has high demand in biotechnological and industrial areas due to its wide range of application. Major applications of cellulases are in textile industry for 'bio-polishing' of clothes, produce stone washed look of denims and also in household detergents to improve fabric softness and brightness¹¹. Cellulase could be used in waste water treatment, the pulp and paper industry and in animal feed¹².

Cellulase is also used in various industries like textile, food, detergent and leather industries⁴. Besides these, which is used in the fermentation of biomass into biofuels¹³, fibre modification and they are even used for pharmaceutical applications. Cellulases are used in effluent treatment, wool and dyeing treatment and in cotton preparations.

The aim of this study is to isolate and screening of cellulase enzyme producing bacteria from the polluted water sources. Identify the organism and produce cellulase enzyme using various growth parameters such as incubation temperature, pH, different concentrations of CMC and incubation time by DNS (dinitro salicylic acid) method to optimize the maximum cellulase production.

MATERIALS AND METHODS

Isolation of organisms

Water samples were collected from polluted water bodies of different places in agricultural fields. Samples were collected in sterile containers and aseptically transfer to the laboratory within an hour. The isolation of organisms was done by Serial dilution total plate count method. Pure cultures of selected colonies were stored in glycerol stock for the further uses.

Screening of cellulose degrading bacteria

Selected colonies were streak plated on CMC agar (Carboxy methyl cellulose -Peptone . 2g, CMC. 2 g , K₂HPO₄ . 4g , Agar .2g , MgSO₄ .0.06g , (NH₂)₂SO₄ . 0.50g , Gelatin .0.4g.) plates to screen for cellulase production by qualitative plate assay. After the incubation period of 24hr at 37°C, CMC agar plates were flooded with 1 % congo red and allowed to stand for 15 min at room temperature and counterstained with 1M NaCl. Clear zone appearing around the growth of bacterial culture indicating positive result of cellulose hydrolysis. The organism able to produce largest clear zone around was used for the following studies.

Identification of an Organism

The selected organism was identified based on morphological, biochemical and physiological characters according to Bergey's manual of determinative bacteriology.

Crude Enzyme Preparation

Pre inoculum was prepared in nutrient broth using *Micrococcus sp* , 200µl of pre inoculum was added in to 100ml of sterile CMC broth medium, Incubate the medium at 37°C for 24hr .1 ml of the broth medium was transferred to micro centrifuge tubes and centrifuged at 4000rpm for 15 min at 4°C and discard the pellet. Supernatant was used as the crude enzyme source which was stored for further enzyme assay.

Optimization of enzyme production

The influence of environmental factors such as pH, temperature, incubation time and concentration of carboxy methyl cellulose in cellulase production were studied to determine the optimum growth conditions for higher production of an enzyme. The parameters checked were, pH range from 6 to 9 (6, 7, 8, 9), incubation time (24, 48, 72hrs) and in 5 different concentrations of CMC (0.5, 1, 1.5, 2 , 2.5%).

Carboxy Methyl Cellulose assay

Cellulase assay parameters like pH, temperature and incubation time were checked separately with 0.5, 1, 1.5, 2 , 2.5% of Carboxymethyl cellulose . Cellulase activity was assayed using dinitro salicylic acid (DNS) reagent¹⁴ by estimation of reducing sugars released from CMC solubilised in 0.05 M phosphate buffer at pH 8. 0.5 ml of Crude enzyme added to 0.5 ml of CMC in 0.05 M phosphate buffer and incubated at 50°C for 30 min. After the incubation time, reaction was stopped by adding 1.5ml of DNS reagent and boiled at 100°C in water bath for 10 min. By measuring absorbance at 540 nm, sugars liberated were determined. One unit (IU) of enzyme activity is expressed as the quantity of enzyme, which is required to release 1µmol of glucose per minute under standard assay conditions.

RESULTS AND DISCUSSION

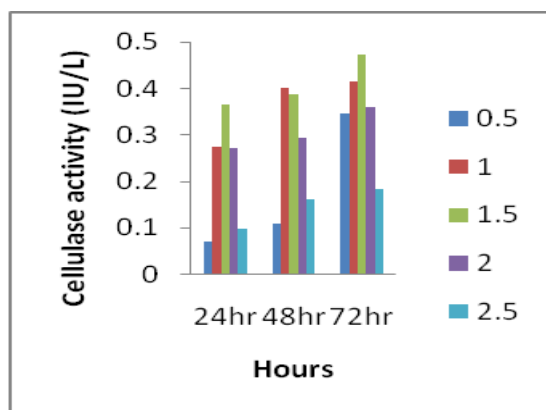
From the result of serial dilution plate count method, 14 different organisms were isolated from the polluted water bodies . All organisms were screened by qualitative plate assay method on Carboxymethyl cellulose and only three organisms were shown zone of clearance around . More clear zone producer was identified as *Micrococcus sp* based on Bergey's manual of determinative bacteriology and used as a test organism. Cellulase enzyme producing property of some bacterial species such as *Cellulomonas sp*, *Pseudomonas sp*, *Bacillus sp* and *Micrococcus sp* were reported¹⁵.

Optimization results shown the higher production of cellulase enzyme was 0.9490 IU/L . pH is the one of the important factor in enzyme production. High acidic and high basic pH values adversely affected enzyme production¹⁶. The pH 6, 7, 8 and 9 were checked in this study and maximum enzyme production was assayed at pH 8 than the other low and high pH. Most of the *Bacillus sp* has an optimum pH 5 for higher production of cellulase¹⁷. Substrate concentration play a major role in production of an enzyme.

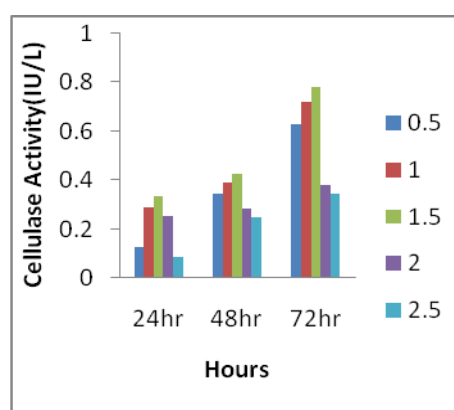
Utilization of CMC as carbon source is best for microbial cellulase production¹⁸. In present, 5 concentrations, 0.5, 1, 1.5, 2 , 2.5% of Carboxymethyl cellulose were used for study to determine the optimum concentration of Carboxymethyl cellulose and higher enzyme activity. The higher enzyme activity was at 1.5% and enzyme produced more at 72hr of incubation time. Lesser activity was shown in 24hr and 48hr of incubation time. Some of *Streptomyces sp*. has been reported to produce higher cellulase enzyme in 72-120 hr of fermentation^{19,20,21}. 37°C was the optimum

temperature for *Micrococcus sp* more enzyme production than 27°C. Optimum temperature for higher cellulase production was 30°C and the lowest yield was achieved at 45 °C²². *Cellulomonas* ASN2 isolated from soil²³ exhibited its optimum cellulase activity at pH of 7.5 and temperature of 60°C²⁴ *Bacillus* strain M9 and NZ showed more enzyme activity at 72 h incubation. From the optimization results, 1.5% of CarboxyMethylCellulose concentration, pH 8, 72 hr of incubation time and 37°C of incubation

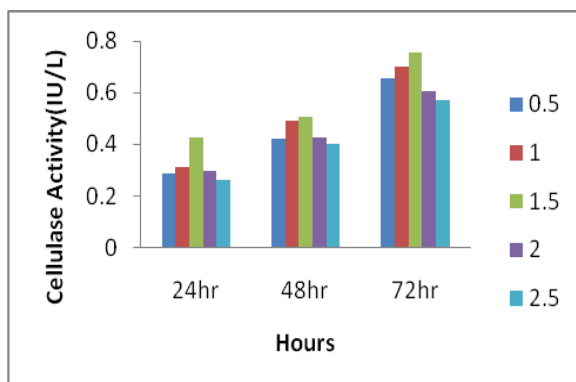
temperature are the optimum growth parameters for maximum production of cellulase enzyme production from *Micrococcus sp*. By following this conditions ,could produce cellulase enzyme easily ,inexpensive production and without risks. **(Figure:-1-Cellulase Activity (IU/L) is shown below).** Optical density of each sample with reaction mixture was taken at 540 nm in a spectrophotometer .



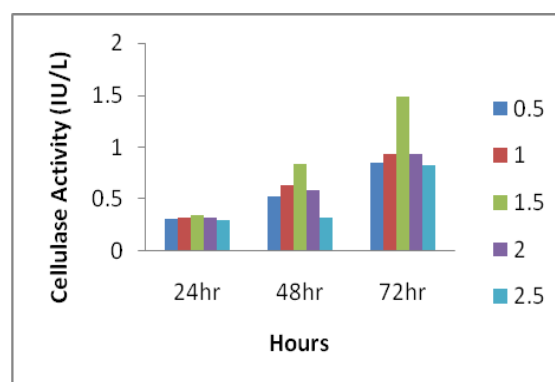
At pH-7, 27°C



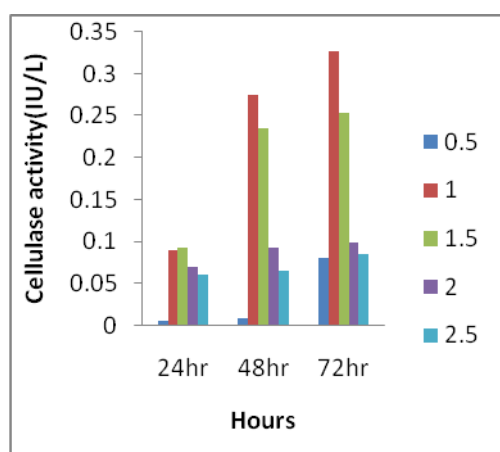
At pH-7, 37°C



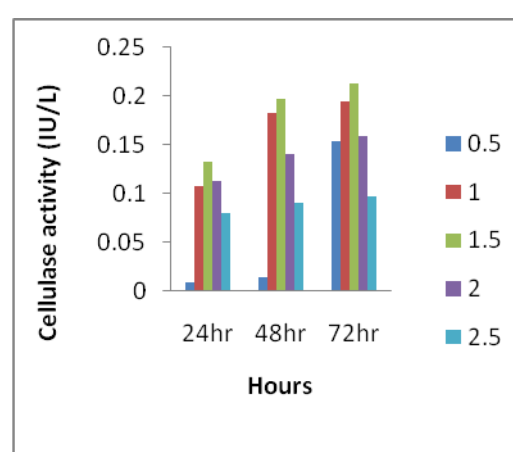
At pH-8, 27°C



At pH-8, 37°C



At pH-6, 27°C



At pH-6, 37°C

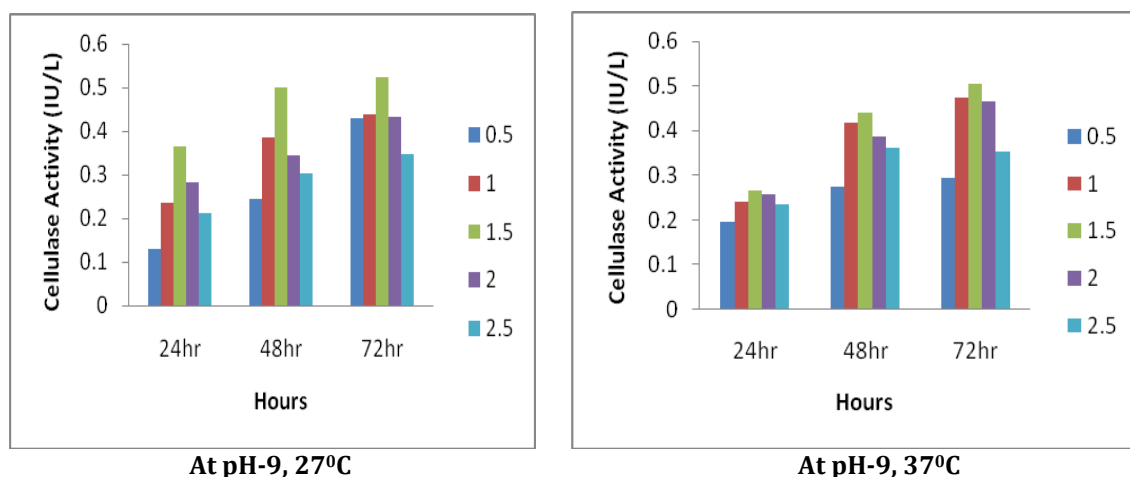


Fig. 1: Cellulase Activity (IU/L)

REFERENCES

- Li YH. A novel thermo acidophilic endoglucanase. Ba- EGA, from a new cellulose degrading bacterium, *Bacillus* sp. AC-1. *Appl Microbiol Biotechnol.* 2006;70:430-436.
- Gao J. Production and characterization of cellulolytic enzymes from the thermo acidophilic fungal *Aspergillus terreus* M11 under solid-state cultivation of corn stover. *Bioresour Technol.* 2008;99:7623-7629.
- Kirk O. Industrial enzyme applications. *Curr Opin Biotechnol.* 2002;13:345-351.
- Bhat MK. Cellulases and related enzymes in biotechnology. *Biotechnol Adv.* 2000;18:355-383.
- Bischoff KM. Purification and characterization of a family 5 endoglucanase from a moderately thermophilic strain of *Bacillus licheniformis*. *Biotechnology Letters.* 2006;28: 1761-1765.
- Camassola M. Characterization of the cellulose complex of *Penicillium echinulatum*, *Biocatalysts Biotransformation.* *AJP.* 2004;22:391-396.
- Haakana H. Cloning of cellulase genes from *Melenocarpus albomyces* and their efficient expression in *Trichoderma reesei*. *Enzyme Microbial Technology.* 2004;34:159-167.
- Doi RH. Cellulase of mesophilic microbes: cellulosome and non-cellulosome producers. *Ann NY Acad Sci.* 2008;1125:267-279.
- Levin L and Forchiassin F. Effect of carbon and nitrogen sources on the cellulolytic activity of *Trametes trogii*. *Rev Argent Microbiol.* 1995;27(1):11-20.
- Immanuel G. Production of endoglucanase by using different nitrogen sources by *Streptomyces* sp. *Int J Microbiol.* 2007;1:24.
- Cavaco-Paulo A. Mechanism of cellulase action in textile processes. *Carbohydr Polym.* 1998;37:273-277.
- Suchita N and Ramesh CK. Bleaching of wheat straw-rich soda pulp with xylanase from a thermoalkalophilic *Streptomyces cyaneus* SN32. *Biores Technol.* 2006 ;97:2291-2295.
- Cherry JR and Fidantsef AL. Directed evolution of industrial enzymes: an update. *Curr Opin Biotechnol.* 2003;14:438-443
- Miller GL. Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Anal Chem.* 1959;31:426-428.
- Nakamura K and Kappamura KJ. Isolation and identification of crystalline cellulose hydrolysing bacterium and its enzymatic properties. *Ferment Technol.* 1982;60(4):343-348.
- Bhat MK and Bhat S. Cellulose degrading enzymes and their potential industrial applications. *Biotechnol Adv.* 1997;15:583-620.
- Haltrich D. Production of fungal xylanases. *Biores Technol.* 1996;58(2):137-161.
- Fukumori F. Purification and properties of a cellulase from alkalophilic *Bacillus*

- sp. Gen. Microbiol. 1985;131(12):3339-3345.
19. Das A. Production of cellulase from a thermophilic *Bacillus* sp isolated from cow dung. *American-Eurasian J Agric Environ Sci.* 2010;8(6):685-691.
 20. Okeke BC and Paterson A. Simultaneous production and induction of cellulolytic and xylanolytic enzymes in a *Streptomyces* sp. *World Journal of Microbiology and Biotechnology.* 1992;8:483-487.
 21. Alam MZ. Isolation, Purification, Characterization of Cellulolytic Enzymes Produced by the Isolate *Streptomyces omiyaensis*. *Pak J BiolSci.* 2004;7:1647-1653.
 22. Harchand RK and Singh S. Characterization of cellulose complex of *Streptomyces albaduncu*. *Journal of Basic Microbiology.* 1997;37(2):93-103.
 23. Muhammad Irfan. Isolation and screening of cellulolytic bacteria from soil and optimization of cellulase production and activity. *Turk J Biochem.* 2012;37(3):120-128.
 24. Sheik Nizamudeen and Bajaj BK. A novel thermo-alkali tolerant endoglucanase production using cost-effective agricultural residues as substrates by a newly isolated *Bacillus* sp. NZ, *Food Technology Biotechnology.* 2009;47(4):435-440.