

BIOCHEMICAL COMPOSITION OF SEAWEEDS AND THEIR ANTI-CANCER PROPERTIES AGAINST HUMAN PAPILLOMAVIRUS (HPV) – A REVIEW

Ashwini S and Manjula Shantaram*

Department of Studies in Biochemistry, Mangalore University, Post Graduate Centre, Chikka Aluvara, 571 232, Kodagu, Karnataka, India.

ABSTRACT

Seaweeds were traditionally used as food stuff in Asian countries for centuries, as early as 2700 BC. Most recently seaweeds have been utilized in Japan as raw materials in the manufacture of many seaweed food products such as jam, cheese, wine, tea, soup and noodles. Seaweeds are highly nutritive foods containing proteins, vitamins, minerals, fiber contents and essential fatty acids. Seaweeds have become a valuable vegetable (fresh or dried) and an important food ingredient in the human diet. Seaweeds constitute some of the most important reservoirs of new therapeutic compounds for humans. Several of them have been shown to have many biological activities, including anticancer activity. Marine algae- derived products play an important role in preventing inflammatory reactions and carcinogenesis by modulating the effects of oxidative stress.

Keywords: Seaweeds, HPV, carbohydrates, proteins, lipids, minerals, vitamins.

INTRODUCTION

Cancer of the cervix uteri is the 4th most common cancer among worldwide with an estimated 527,624 new cases and ranks as the 2nd cause of female cancer in India with 265,653 deaths in 2012 and 122,844 new cervical cancer cases diagnosed annually in India¹. Virtually 100% of cervical tumors are attributable to HPV infection which is annually generating 530,000 cervical cancer cases worldwide^{2,3}. There are highly effective prophylactic HPV vaccines for HPV infection which are publicly available for preventing the transmission of HPV in the developed countries.

However, there are certain drawbacks for these vaccines that they are relatively expensive and are likely to be Papillomavirus restricted in their protection⁴. Thus, these vaccines may not be affordable and initially not available to women in all parts of the world. Further, they may not offer protection against all cancers associated with different HPV types. Therefore, the urge in search for potential drug candidates against various HPV strains which has higher inhibitory activities is increasing in the pharmaceutical industry. With this regard, natural bioactive compounds derived from marine sources are found to have great sources for the development of new generation anti-HPV therapeutics, which

is more effective with a fewer side effects. Among which Seaweeds or marine algae constitute one of the commercially important renewable marine living resources. Seaweeds have been an important dietary component since, at least fourth century in Japan and sixth century in China. In Korea, consumption of seaweeds is a common practice⁵.

Based on their pigmentation, seaweeds are classified into Chlorophyta (Green algae), Rhodophyta (Red algae) and Phaeophyta (Brown algae). Seaweeds such as *Caulerpa*, *Ulva*, *Enteromorpha*, *Codium* and *Monostroma* (Green algae; Figure 1); *Sargassum*, *Hydroclathrus*, *Padina*, *Macrocystis* (Brown algae; Figure 2); *Porphyra*, *Gracilaria*, *Eucheuma*, *Laurencia* and *Acanthophora* (Red algae; Figure 3) are used for human consumption in the form of soup, salad, curry which are protein rich. Marine algae are also used in different parts of the world as animal feed and fertilizer, they contain more than 60 trace elements, carbohydrate, iodine, bromine, vitamins and some bioactive substances⁶.

Certain seaweeds have been used for the treatment of cancer, many crude or partially purified polysaccharides from various brown, green, and red algae have been tested for their antitumor activities⁷. Fucoidan isolated from

brown seaweed such as *Undaria* and *Laminaria* showed anticoagulant, antiviral and anticancer properties^{8,9}. Seaweeds have become a valuable vegetable (fresh or dried) and an important food ingredient in the human diet¹⁰.

Several studies have reported that compounds extracted from seaweed may be effective anticancer agents. This review will mainly focus on the biochemical compounds present in marine organisms and the importance of seaweed, their potential as therapeutic application against cervical cancer.

Biochemical composition of seaweeds

The protein, carbohydrate and organic carbon content was estimated in 43 marine algal species collected from different marine stations along the Maharashtra coast among which more protein and carbohydrate content was recorded in Chlorophycean, Rhodophycean and Phaeophycean algae by Dhargalkar et al.,¹¹.

Chakraborty and Santra recorded higher carbohydrate in the green seaweeds *U. lactuca* (35.27%) and *Enteromorpha intestinalis* (30.58%)¹². The carbohydrate content of seaweed collected from Mandapam coast varied from 20.47 to 23.9% and maximum carbohydrate concentration was recorded from brown algae *Turbinaria conoides* (23.9%), *S. tenerimum* (23.55%), *S. wightii* (23.50%), followed by the green alga *E.intestinalis* (23.84%), and red algae *Hypnea valentiae*(23.60%), *Acanthophora spicifera* (23.54%). While the minimum carbohydrate content was reported from green alga *Codium tomentosum*(20.47%) followed by brown algae *Padina gymnospora* (21.88%), *Colpomenia sinuosa* (22.46%) and the red alga *G. folifera* (22.32%)¹³.

Fifty individual plants of *Saccharina*, *Fucus* (*serratus* and *spiralis*) and *Ascophyllum* were recorded for glucose content and were found to have 65%, 30% and 20% of the total sugars in an autumn sample respectively¹⁴. The study percentage of *Hypnea valentiae* for carbohydrate content was higher than those of several seaweed species of the red algae genus *Hypnea* collected in Darwin Harbour¹⁵.

Concentration of total polysaccharides in the seaweed species of interest, range from 4% to 76% of their dry weight; among which highest contents are found in species such as *Ascophyllum*, *Porphyra* and *Palmaria* (Table1). Green seaweed species such as *Ulva* also have a high polysaccharide content, up to 65% of dry weight. Arasaki and Arasaki observed seaweeds for their richness in polysaccharides, minerals and certain vitamins¹⁶.

Brown alga *Tubinaria ornata* from Gulf of Mannar region was observed for highest protein

content by Dinesh et al.,¹⁷. Mairh et al.,¹⁸ reported 22.22% of crude protein in *Ulva fasciata*, which is frequently consumed under the name of "ao-nori" by the Japanese people, was observed for high protein level between 20 and 26% (dry product)¹⁹. Red seaweeds such as *Porphyra tenera* (47% of dry mass) or *Palmaria palmata* (35% of dry mass) were recorded higher protein levels (Table 2)²⁰.

The major metabolites such as proteins, carbohydrates and lipids were estimated by Dhargalkar²¹. Red algae (Rhodophyceae) were collected from Mandapam coastal regions southeast coast of India for proximate composition evaluation and the protein content varied from 3.25±0.36 to 17.08±0.28%; maximum protein was recorded in *P. gymnospora* (17.08±0.28%) followed by *E. intestinalis* (16.38±0.50%) and *S. tenerimum*(12.42±0.63%).

Munda reported that in *Fucus* sp. (brown seaweeds), aspartic and glutamic acids can represent between 22 and 44% of the total amino acids²². Marine macro algae varieties contained low amount of lipids; they are the sources of poly unsaturated fatty acids (PUFA). The distribution of fatty acid in seaweed products showed high level of omega-3 fatty acids and demonstrated a nutritionally ideal omega-6/omega-3 free fatty acid ratio. Seaweeds contain up to 2% of dry weight of lipids and much of this lipid content is made up of polyunsaturated fatty acids^{23,24}.

The lipid content of seaweeds varied from 1.33 to 4.6; in that the maximum lipid content was observed in *Enteromorpha clathrata* (4.6%; Figure 1) followed by *G. folifera* (3.23%), *Codium tomentosum* (2.53%), *C. sinuosa* (2.337%) and *S. wightii* (2.337%). The minimum lipid concentration was observed in *E. intestinalis* (1.33%) followed by *P. gymnospora* (1.4%), *S. tenerimum* (1.46%) and *U. lactuca* (1.6%)¹³. In Japan, the lipid content of *P. gymnospora* products was exhibited: Biochemical and nutritional aspects (1.4±0.30%), *S. tenerimum* (1.46±0.20%) and *U. lactuca* (1.6±0.17%) values were smaller than those obtained for most of the seaweeds, which range from 2.80±0.23% to 3.49±0.28 %. This value which is relatively low; is comparable to results obtained from previous studies²⁵. The higher result for carbohydrate content (64.00% dry weight.) in *Caulerpa lentillifera* was reported by Nguyen et al.²⁶.

Seaweeds are a well-known source of minerals. An adequate intake of minerals is required for a high nutritional quality of the diet which can help prevention of chronic nutrition-related diseases and degenerative diseases including cancer, cardiovascular disease, Alzheimer's disease, and premature aging^{27,28}. Seaweeds are

considered as a potential material for the production of different nutraceuticals and food supplements^{29,30}. The selected micro nutrients (Fe + Zn + Mn + Cu⁺) of *Panax vietnamensis* were found to be higher (45.5– 309 mg/100 g dry weight) than any of the land vegetables as well as edible seaweeds like *C. lentillifera*, *Enteromorpha flexuosa*, *Monostroma oxysperum*, *Eucheuma denticulatum* and *Gracilaria parvispora* reported from Hawaii³¹.

The mineral content of certain seaweeds was recorded higher than that of land plants and animal products³². Copper is present at a high level in seaweeds³³. Daily intake of 8g of 'Kombu', as used in Asian cooking, contains 65% of the RNI for magnesium³⁴. Seaweeds are described as an ideal food-safe natural source of the mineral³⁵. It is found that 8 g of *Porphyra umbilicalis* (Nori) provides 9 mg of vitamin C, or 15% of the RNI³⁶. *U. lactuca* can provide vitamin B12 in excess of the recommended dietary allowances for Ireland of 1.4 g/day with 5g in 8g of dry foodstuff³⁷. Seaweeds are known as an excellent source of vitamins and minerals³⁸. Chapman and Chapman reported that 100 g of seaweed provides more than the daily requirement of Vitamin A, B2 and B12 and two thirds of the Vitamin C requirement (Table 3)³⁹.

Role of seaweeds in cervical cancer

The vital role of HPV16 in carcinogenesis was considered^{40,41}. Thus most of the efforts for developing therapeutic HPV vaccines has been directed towards development of vaccines against HPV16 viral oncogenes *E6* and *E7*^{42,43}. Though there is still no FDA approved anti-HPV drug listed so far, some Interferons (IFNs) are the only antiviral drugs approved for the therapy of benign HPV related lesions. Chinese medicine *chaihu* possesses good anti-HPV activities; it has inhibitory effects by interfering with the expression of HPV-DNA in genital warts⁴⁴.

Brown seaweed with low molecular weight fucoïdan, mediated the broad-spectrum growth inhibition of human carcinoma cells, including HeLa cervix adenocarcinoma, HT1080 fibrosarcoma, K562 leukemia, U937 lymphoma, A549 lung adenocarcinoma and HL-60⁴⁵. Fucoïdan produced from brown algae was reported to inhibit HPV pseudovirus infection *in vitro* with the IC50 value of 1.1 µg/mL⁴⁶. Brown algae could be a relevant source of anticancer compound⁴⁷. Heterofucans from *Sargassum filipendula* exhibited anti-proliferative effects on cervical cells⁴⁸. Fucoxanthin has been shown to induce apoptosis in human cervical cancer HeLa cells^{49,50}.

Marine polysaccharide carrageenan was able to generate antigen-specific immune responses and anti-tumor effects in female (C57BL/6) mice vaccinated with HPV16 E7 peptide vaccine⁵¹. Brown seaweeds were isolated for active sulfated homo-heterofucans which have shown effective antitumor activities with a wide range of mechanisms which stands as valuable source⁵². Sulfated polysaccharides, such as heparin, cellulose sulfate and dextran sulfate, were reported to block the infectivity of papillomaviruses⁵³. Polysaccharide fraction SF isolated from the brown seaweed *Sargassum stenophyllum* with major fucose was reported to promote morphological modifications in HeLa cells at low (2.5µg/mL) concentrations⁵⁴. Polysaccharides and terpenoids from brown algae are considered as promising bioactive molecules with anticancer activity by Taskin *et al.*⁵⁵, Devery *et al.*,⁵⁶. Polysaccharide-rich extract from *Sargassum filipendula*, *C. Agardh* showed anti-proliferative effect on HeLa cell (human uterine adenocarcinoma cell) proliferation (Table 4)⁵⁷.

Several heparan sulfate proteoglycans (HSPGs) serve as a primary receptor protein in natural HPV infection of keratinocytes as HPV receptors and support a putative role for syndecan⁵⁸. Marine heparinoid polysaccharides such as alginic acid and fucoïdan were reported to effectively block HPV pseudovirion infection just like heparin⁵⁹.

Carrageenans from marine sources were observed to inhibit HPV infection. Gliotoxin isolated from marine fungus *Aspergillus* sp. was found to induce apoptosis in HPV related cancer cells via the mitochondrial pathway leading to cell death⁶⁰. Marine algae are important sources of non-animal sulfated polysaccharides and these biomolecules are widely studied on therapeutic applications such as anti-thrombotic, anticoagulant, antioxidant, anti-inflammatory and anti-proliferative compounds⁶¹⁻⁶⁴.

CONCLUSION

Edible seaweeds have been shown to be high in essential vitamins and minerals that would inflate a balanced diet if consumed regularly. Marine algae are known for natural richness in minerals, vitamins, polyunsaturated fatty acids and their low content lipids as well as high content of bioactive molecules. They can serve as good source of healthy food. Also seaweeds may solve the problems of deficiency of protein, carbohydrate and mineral deficiency in human nutrition by consuming them in daily life. Further seaweed will lead as a novel candidate in pharmaceuticals to develop a natural compound as an anticancer agent for production

of potential anticancer drug and it is necessary to revitalize the use of seaweed in the newly health-conscious consumer environments of several countries.

ACKNOWLEDGEMENT

We are grateful to Mangalore University for the support and encouragement.

CONFLICTS OF INTEREST

No conflict of interest.



Fig. 1: *Enteromorpha clathrata* (Green algae)



Fig. 2: *Padina boerengsnii* (Brown algae)



Fig. 3: *Gracilaria Spp* (Red algae)

Table 1: Seaweed carbohydrate content by 8 g portion compared with guideline daily amounts (GDA) of fiber

| Seaweed | Total fiber* | Soluble fiber* | Insoluble fiber* | Carbohydrates* | Fiber as % GDA [†] |
|-----------------------------|--------------|----------------|------------------|----------------|-----------------------------|
| <i>Ascophyllum nodosum</i> | 2.8 | 2.4 | 0.4 | 4.2 | 11.6 |
| <i>Laminaria digitata</i> | 3.0 | 2.6 | 0.4 | 4.8 | 12.5 |
| <i>Himanthalia elongate</i> | 2.6 | 2.1 | 0.6 | 4.0 | 10.8 |
| <i>Undaria pinnatifida</i> | 2.8 | 2.4 | 0.4 | 3.9 | 11.6 |
| <i>Porphyra umbilicalis</i> | 2.7 | 2.1 | 0.7 | 3.8 | 11.25 |
| <i>Palmaria palmata</i> | 2.7 | 1.5 | 1.2 | 5.3 | 11.25 |
| <i>Ulva spp.</i> | 3.0 | 1.7 | 1.3 | 3.3 | 12.5 |
| <i>Enteromorpha spp.</i> | 3.0 | 1.8 | 1.3 | 4.8 | 12.5 |

*Values from the Institut de Phytonutrition (2004).³⁴

[†]Guideline daily amounts from the Institute of Grocery Distribution (2006).⁶⁸

Table 2: Composition of amino acids of some seaweeds (in g amino acid/100 g protein)

| Amino acids | <i>Ulva armoricana</i> ¹⁰ (green seaweed) | <i>Ulva Pertusa</i> ¹⁹ (green seaweed) | <i>Palmaria palmata</i> ⁶⁹ (red seaweed) | <i>Porphyra tenra</i> ¹⁹ (red seaweed) |
|---------------|---|--|--|--|
| Histidine | 1.2±2.1 | 4.0 | 0.5±1.2 | 1.4 |
| Isoleucine | 2.3±3.6 | 3.5 | 3.5±3.7 | 4.0 |
| Leucine | 4.6±6.7 | 6.9 | 5.9±7.1 | 8.7 |
| Lysine | 3.5±4.4 | 4.5 | 2.7±5.0 | 4.5 |
| Methionine | 1.4±2.6 | 1.6 | 2.7±4.5 | 1.1 |
| Phenylalanine | 5.0±7.1 | 3.9 | 4.4±5.3 | 3.9 |
| Threonine | 4.5±6.8 | 3.1 | 3.6±4.1 | 4.0 |
| Tryptophan | ∅ | 0.3 | 3.0 | 1.3 |
| Valine | 4.0±5.2 | 4.9 | 5.1±6.9 | 6.4 |
| Alanine | 5.5±7.0 | 6.1 | 6.3±6.7 | 7.4 |
| Arginine | 4.3±8.7 | 14.9 | 4.6±5.1 | 16.4 |
| Aspartic acid | 6.0±11.8 | 6.5 | 8.5±18.5 | 7.0 |
| Glutamic acid | 11.7±23.4 | 6.9 | 6.7±9.9 | 7.2 |
| Cysteine | ∅ | ∅ | ∅ | ∅ |
| Glycine | 6.3±7.5 | 5.2 | 4.9±13.3 | 7.2 |
| Proline | 5.0±10.5 | 4.0 | 1.8±4.4 | 6.4 |
| Serine | 5.6±6.1 | 3.0 | 4.0±6.2 | 2.9 |
| Tyrosine | 4.4±4.7 | 1.4 | 1.3±3.4 | 2.4 |

Table 3: Mineral compositions of seaweeds compared to whole foods

| Food Type | Calcium | Potassium | Magnesium | Sodium | Copper | Iron | Iodine | Zinc |
|-------------------------------|---------|-----------|-----------|--------|--------|------|--------|------|
| Seaweed | | | | | | | | |
| (mg/100 g wet weight)* | | | | | | | | |
| <i>Ascophyllum nodosum</i> | 575.0 | 765.0 | 225.0 | 1173.8 | 0.8 | 14.9 | 18.2 | NA |
| <i>Laminaria digitata</i> | 364.7 | 2013.2 | 403.5 | 624.6 | 0.3 | 45.6 | 70.0 | 1.6 |
| <i>Himanthalia elongata</i> | 30.0 | 1351.4 | 90.1 | 600.6 | 0.1 | 5.0 | 10.7 | 1.7 |
| <i>Undaria pinnatifida</i> | 112.3 | 62.4 | 78.7 | 448.7 | 0.2 | 3.9 | 3.9 | 0.3 |
| <i>Porphyra umbilicalis</i> | 34.2 | 302.2 | 108.3 | 119.7 | 0.1 | 5.2 | 1.3 | 0.7 |
| <i>Palmaria palmata</i> | 148.8 | 1169.6 | 97.6 | 255.2 | 0.4 | 12.8 | 10.2 | 0.3 |
| <i>Chondrus crispus</i> | 373.8 | 827.5 | 573.8 | 1572.5 | 0.1 | 6.6 | 6.1 | NA |
| <i>Ulva spp.</i> | 325.0 | 245.0 | 465.0 | 340.0 | 0.3 | 15.3 | 1.6 | 0.9 |
| <i>Enteromorpha spp.</i> | 104.0 | 351.1 | 455.1 | 52.0 | 0.1 | 22.2 | 97.9 | 1.2 |
| Whole food | | | | | | | | |
| (mg/100 g weight)† | | | | | | | | |
| Brown rice | 110.0 | 1160.0 | 520.0 | 28.0 | 1.3 | 12.9 | NA | 16.2 |
| Whole milk | 115.0 | 140.0 | 11.0 | 55.0 | Tr | 0.1 | 15.0 | 0.4 |
| Cheddar cheese | 720.0 | 77.0 | 25.0 | 670.0 | 0.0 | 0.3 | 39.0 | 2.3 |
| Sirloin steak | 9.0 | 260.0 | 16.0 | 49.0 | 0.1 | 1.6 | 6.0 | 3.1 |
| Lentils green and brown | 71.0 | 940.0 | 110.0 | 12.0 | 1.0 | 11.1 | NA | 3.9 |
| Spinach | 170.0 | 500.0 | 54.0 | 140.0 | 0.0 | 2.1 | 2.0 | 0.7 |
| Bananas | 6.0 | 400.0 | 34.0 | 1.0 | 0.1 | 0.3 | 8.0 | 0.2 |
| Brazil nut | 170.0 | 660.0 | 410.0 | 3.0 | 1.8 | 2.5 | 20.0 | 4.2 |
| Peanuts | 60.0 | 670.0 | 210.0 | 2.0 | 1.0 | 2.5 | 20.0 | 3.5 |

*Values for seaweeds from the Institut de Phytonutrition (2004).³⁴†Values for whole foods from McCance et al. (1993).⁶⁷**Table 4: Anti-HPV and related anticancer agents from seaweeds**

| Marine Organisms | Specific Compounds | Mechanisms of Action |
|------------------|-------------------------------|--|
| Red Algae | λ-carrageenan ^{4,65} | Blocking HPV infection |
| | κ-carrageenan ^{4,65} | Blocking HPV infection |
| | ι-carrageenan ^{4,65} | Blocking HPV infection |
| | Agar ⁴ | Blocking HPV infection |
| Brown Algae | Alginate acid ^{4,66} | Inhibiting HPV and cancer cell proliferation |
| | Fucoidan ⁴ | Blocking HPV infection |

REFERENCES

1. Bruni L, Barrionuevo-Rosas L, Albero G, Aldea M, Serrano B, Valencia S, Brotons M, Mena M, Cosano R, Muñoz J, Bosch FX, de Sanjosé S and Castellsagué X. ICO Information Centre on HPV and Cancer (HPV Information Centre). Human Papillomavirus and Related Diseases in India. Summary Report 2015-03-20. [Data Accessed].
2. Forman, D, Martel C, Lacey CJ, Soerjomataram I, Lortet TJ, Bruni L, Vignat J, Ferlay J, Bray F and Plummer M. Global burden of human papillomavirus and related diseases. *Vaccine*. 2012;30:12-23.
3. Lowy DR and Schiller JT. Reducing HPV-associated cancer globally. *Cancer Prev Res*. 2012;5:18-23.
4. Buck CB, Thompson CD, Roberts JN, Muller M, Lowy DR and Schiller JT. Carrageenan is a potent inhibitor of papillomavirus infection. *PLoS Pathog*. 2006;2:69.
5. Venugopal V. marine products for health, functional food and nutritional series. CRC press, NY. 2009;261 – 272.
6. Silas EGVSK. Chennubhotla and N. Kaliaperumal. Seaweed resources, products and utilisation *Seaweed Res. Uti In* 1986;9(1&2): 11-24.
7. Ramberg JE, Nelson ED and Sinnott RA. Immunomodulatory dietary polysaccharides: A systematic review of the literature *Nut J*. 2010;9: 54.
8. Chevolut L, Mulloy B, Ratiskol J, Foucault A and Collic-Jouault S. A disaccharide repeat unit is the major structure in fucoidans from two species of brown algae. *Carbohydr Res*. 2001;330:529-535.
9. Zhuang Cun ZC, Itoh H, Mizuno T and Ito H. Antitumor active fucoidan from the brown seaweed, *Umitoranoo* (*Sargassum thunbergii*). *Bioscience, Biotechnology and Biochemistry*. 1995;59(4): 563-567.
10. Fleurence J. Seaweed proteins: Biochemical, nutritional aspects and potential uses. *Trends in Food Science and Technology*. 1999;10: 25-28.
11. Dhargalkar VK, Jagtap TG and Untawale AG. Biochemical constituents of seaweeds along the Maharashtra coast. *Indian J Mar Sci*. 1980;2(4):297-299.
12. Chakraborty S and Santra SC. Biochemical Composition of Eight Benthic Algae Collected from Sunderban. *Indian J Mar Sci*. 2008;37(3):329-332.
13. Park EJ and Pezzuto JM. Antioxidant Marine Products in Cancer Chemoprevention. *Antioxid Redox Signal*. 2013; 19: 115-138.
14. Jensen A. Component sugars of some common brown algae, Akademisk Trykningsentral, Blindern, Oslo. Norwegian Institute of Seaweed Research. 1956; Report 9: 1-8.
15. Renaud SM, Lambridinis G, Luong-Van T, Parry DL and Lee C. Chemical composition of algae for use in *Trochus niloticus* studies. In: L. Lee and P. W Lynch, Eds.), *Trochus: Status, Hatchery Practice and Nutrition*, Australian Centre for International Agricultural Research, Canberra, 1997; 88-96.
16. Arasaki A and Arasaki T. Low calories, High Nutrition. *Vegetables from the Sea to Help you Look and Feel Better*, Japan Publications Inc. 1983;39-42.
17. Dinesh G, Sekar M and Kannan R. Nutritive properties of seaweeds of Gulf of Mannar, Tamil Nadu. *Seaweed Res Utiln*. 2007; 29(1&2): 125-132.
18. Mairh OP, Ohno M and Matsuoka M. Culture of brown alga *Laminaria japonica* (Phaeophyta, Laminariales) in warm waters of Shikoku, Japan. *Indian J Mar Sci*. 1991;20:55-60.
19. Fujiwara-Arasaki T, Mino N and Kuroda M. The protein value in human nutrition of edible marine algae in Japan' in *Hydrobiologia*. 1984;116/117: 513-516.
20. Morgan KC, Wright JLC and Simpson FJ. Review of chemical constituents of the red alga *Palmaria palmata* (Dulse) in *Econ Bot*. 1980;34: 27-50.
21. Dhargalkar VK. Biochemical studies on *Ulva reticulata* Forsskal. *Proceeding International Symposium on Marine Algae of the Indian Ocean Region*, CSMCRI, Bhavnagar. 1979;40.
22. Munda IM. 'Differences in amino acid composition of estuarine and marine fucoids' in *Aquatic Botany*. 1977; 3: 273-280.
23. Marsham S, Scott GW and Tobin ML. Comparison of nutritive chemistry of a range of temperate seaweeds. *Food Chem*. 2007;100:1331-1336.
24. Sanchez-Machado DI, Lopez-Hernandez J, Paseiro-Losada P and Lopez-Cervantes J. Fatty acids, total lipid, protein and ash contents of processed edible seaweeds. *Food Chem*. 2004;85:439-444.
25. Integaard M and Minsaas J. In animal and human nutrition. *Seaweed*

- Resources in Europe. Uses and Potential (Guiry, M.D. and G. Blunden, Eds.) John Wiley and Sons. 1991;21-64.
26. Nguyen VT, Ueng JP and Tsai GJ. Proximate Composition, Total Phenolic Content, and Antioxidant Activity of Seagrass (*Caulerpa lentillifera*). *Journal of Food Science*. 2011;76(7):950-958.
 27. Fenech M and Ferguson LR. Vitamins/minerals and genomic stability in humans. *Mutat Res*. 2001;475:1-6.
 28. Kersting M, Alexy U and Sichert-Hellert W. Dietary intake and food sources of minerals in 1 to 18 year old German children and adolescents. *Nutr Res*. 2001;21:607-616.
 29. Martínez-Navarrete N, Camacho MM, Martínez-Lahuerta J, Martínez-Monzo J and Fito P. Iron deficiency and iron fortified foods—A review. *Food Res Int*. 2002;35:225-231.
 30. Shahidi F. Nutraceuticals and functional foods: Whole versus processed foods. *Trends Food Sci Technol*. 2009;20:376-387.
 31. McDermid KJ and Stuercke B. Nutritional composition of edible Hawaiian seaweeds. *Journal of Applied Phycology*. 2003;15:513-524.
 32. Ito K and Hori K. Seaweed: chemical composition and potential uses. *Food Review International*. 1989;5:101-144.
 33. EU Health and Consumer Directorate-General. Council directive on nutrition labeling for foodstuffs 90/496/ EC. *Official Journal of the European Communities*. No L 276/40. 1990.available at http://ec.europa.eu/food/food/labellingnutrition/nutritionlabel/index_en.htm. Accessed: October 25, 2007.
 34. Institut de Phytonutrition. Functional, health and therapeutic effects of algae and seaweed. Beausoleil, France, Institut de Phytonutrition electronic database. Version 1.5. Institut de Phytonutrition. 2004.
 35. Teas J. Dietary brown seaweeds and human health effects. *Advances in applied phycology utilisation*. In: Critchley AT, Ohno M, Largo DB. eds. *World Seaweed Resources*. Amsterdam, ETI Bioinformatics; Section 9. 2006.
 36. Committee on Medical Aspects of Food and Nutrition Policy. *Dietary Reference Values for Food Energy and Nutrients for the United Kingdom*. Rep Health Soc Subj. 1991;41:1-210.
 37. Food Safety Authority of Ireland. *Recommended Dietary Allowances for Ireland*. Dublin, Food Safety Authority of Ireland. 1999.
 38. Lahaye M. Marine algae as sources of fibers: determination of soluble and insoluble dietary fiber contents in some sea vegetables. *Journal Science of Food Agricultural*. 1991;54:587-594.
 39. Chapman VJ and Chapman DJ. (Eds.). *Seaweeds and their uses* New York: Chapman & Hall. 3rd ed., 1980;25-42.
 40. Zur HH. Papillomaviruses and cancer: From basic studies to clinical application. *Nat. Rev. Cancer* 2002;2:342-350.
 41. Steenbergen RD, De WJ, Wilting SM, Brink AA, Snijders PJ and Meijer CJ. HPV-mediated transformation of the anogenital tract. *J Clin Virol*. 2005;32:25-33.
 42. Borysiewicz LK, Fiander A, Nimako M, Man S, Wilkinson GW and Westmoreland . A recombinant vaccinia virus encoding human papillomavirus types 16 and 18, E6 and E7 proteins as immunotherapy for cervical cancer. *Lancet*. 1996;347:1523-1527.
 43. Peng S, Trimble C, Alvarez RD, Huh WK, Lin Z and Monie A. Cluster intradermal DNA vaccination rapidly induces E7-specific CD8+ T-cell immune responses leading to therapeutic antitumor effects. *Gene Ther*. 2008;15:1156-1166.
 44. Li J, Luo KZ, Lin Y, Chen WN and Lai SP. Laboratory study on anti-human papillomavirus activity of Bupleurum chinese. *Chin J Dermatol Venerol Integ Trad W Med*. (In Chinese). 2005;4:171-173.
 45. Zhang Z, Teruya K, Eto H and Shirahata S. Fucoïdan extract induces apoptosis in MCF-7 cells via a mechanism involving the ROS-dependent JNK activation and mitochondria-mediated pathways. *1371/journal.pone.0027441* (2011) 1-14.
 46. Buck CB, Thompson CD, Roberts JN, Muller M, Lowy DR and Schiller JT. Carrageenan is a potent inhibitor of papillomavirus infection. *PLoS Pathog*. 2006;2:69.
 47. Manivannan K, Thirumaran G, Karthikai Devi G, Hemalatha A and Anantharaman P. Biochemical Composition of Seaweeds from Mandapam Coastal Regions along Southeast Coast of India, *American-Eurasian J of Bot*. 2008;1(2): 32-37.

48. Costa LS, Fidelis GP, Telles CB, Dantas-Santos N, Camara RB, Cordeiro SL, Costa MS, Almeida-Lima J, Melo-Silveira RF and Oliveira RM. Antioxidant and antiproliferative activities of heterofucans from the seaweed *Sargassum filipendula*. *Mar Drugs*. 2011;9(6):952-966.
49. Hou L, Gao C, Chen L, Hu G and Xie S. Essential role of autophagy in fucoxanthin-induced cytotoxicity to human epithelial cervical cancer HeLa cells. *Acta Pharmacol Sin*. 2013;34:1403-1410.
50. Ye G, Lu Q, Zhao W.; Du, D.; Jin, L.; Liu, Y. Fucoxanthin induces apoptosis in human cervical cancer cell line HeLa via PI3K/Akt pathway. *Tumour Biol*. 2014;35:11261-11267.
51. Zhang YQ, Tsai YC, Monie A, Hung CF and Wu TC. Carrageenan as an adjuvant to enhance peptide-based vaccine potency. *Vaccine*. 2010;28:5212-5219.
52. Xin XL, Geng MY, Guan HS and Li ZL. Study on the mechanism of inhibitory action of 911 on replication of HIV-1 in vitro. *Chin J Mar Drugs*. 2000;19:15-18.
53. Bousarghin L, Touzé A, Sizaret PY and Coursaget P. Human papillomavirus types 16, 31, and 58 use different endocytosis pathways to enter cells. *J Virol*. 2003;77:3846-3850.
54. Stevan FR, Oliveira MB, Bucchi DF, Nosedá, Iacomini M and Duarte ME. Cytotoxic effects against HeLa cells of polysaccharides from seaweeds. *J Submicrosc Cytol Pathol*. 2001;33:477-484.
55. Taskin E, Caki Z, Ozturk M and Taskin E. Assessment of in vitro antitumoral and antimicrobial activities of marine algae harvested from the eastern Mediterranean sea. *Afr J Biotechnol*. 2010; 9: 4272-4277.
56. Devery R, Miller A and Stanton C. Conjugated linoleic acid and oxidative behavior in cancer cells. *Biochem Soc Transact*. 2001; 29: 341- 344.
57. Costa LS, Fidelis GP, Cordeiro SL, Oliveira RM, Sabry DA, Câmara RBG, Nobre LTDB, Costa MSSP, Jailma AL and Farias EHC. Biological activities of sulfated polysaccharides from tropical seaweeds. *Biomed Pharmacother*. 2010;64:21-28.
58. Saeed SK, Alessandra H, Ernst K, Guerrino M, Katharina S and Reinhard K. Different heparin sulfate proteoglycans serve as cellular receptors for human papillomaviruses. *J Virol*. 2003;77: 13125-13135.
59. Isuru W, Li YX, Vo TS, Quang VT, Ngo DH and Kim SK. Induction of apoptosis in human cervical carcinoma HeLa cells by neoechinulin A from marine-derived fungus *Microsporium* sp. *Process Biochem*. 2013;48:68-72.
60. Nguyen VT, Lee JS, Qian ZJ, Li YX, Kim KN, Heo SJ, Jeon YJ, Park WS, Choi IW and Je JY. Gliotoxin isolated from marine fungus *Aspergillus* sp. induces apoptosis of human cervical cancer and chondrosarcoma cells. *Mar Drugs*. 2014;12:69-87.
61. Barroso EM, Costa LS, Medeiros VP, Cordeiro SL, Costa MS, Franco CR, Nader HB, Leite EL and Rocha HAO. A non-anticoagulant heterofucan has antithrombotic activity in vivo. *Planta Med*. 2008;74:712-718.
62. Cumashi A, Ushakova NA, Preobrazhenskaya ME, D'Incecco A, Piccoli A, Totani L, Tinari N, Morozevich GE, Berman AE, Bilan MI, Usov AI, Ustyuzhanina NE, Grachev AA, Sanderson CJ, Kelly M, Rabinovich GA, Iacobelli S and Nifantiev NE. A comparative study of the anti-inflammatory, anticoagulant, antiangiogenic, and antiadhesive activities of nine different fucoidans from brown seaweeds. *Glycobiology*. 2007;17:541-552.
63. Bilan MI and Usov AI. Structural analysis of fucoidans. *Nat. Prod. Commun*. 2008;3:1639-1648.
64. Jiao G, Yu G, Zhang J and Ewart HS. Chemical structures and bioactivities of sulfated polysaccharides from marine algae. *Mar Drugs*. 2011;9:196-223.
65. Roberts JN, Kines RC, Katki HA, Lowy DR and Schiller JT. Effect of Pap smear collection and carrageenan on cervicovaginal human papillomavirus-16 infection in a rhesus macaque model. *J. Natl. Cancer Inst*. 2011;103:737-743.
66. Stevan FR, Oliveira MB, Bucchi DF, Nosedá, Iacomini M and Duarte M.E. Cytotoxic effects against HeLa cells of polysaccharides from seaweeds. *J. Submicrosc. Cytol Pathol*. 2001;33:477-484.
67. McCance RA, Widdowson EM and Holland B. McCance and Widdowson's Composition of Foods. Cambridge, Royal Society of Chemistry; 6th ed. 1993.
68. Institute of Grocery Distribution. Best Practice Guideline on the Presentation of Guideline Daily Amounts. London,

Institute of Grocery Distribution; 2006. Available at: <http://www.igd.com/CIR.asp? menuid 36 and cirid 1877>. Accessed: October 25, 2007.

69. Indegaard M and Minsaas J. In Animal and Human Nutrition. Seaweed Resources in Europe. Uses and Potential. Guiry, M.D. and Blunden, G., eds, John Wiley and Sons. 1991;21-64.